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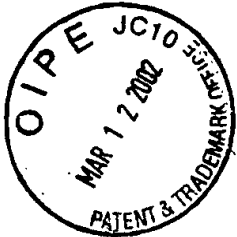
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MEANS FOR IDENTIFYING THE LOCUS OF A MAJOR RESISTANCE  
GENE TO THE RICE YELLOW MOTTLE VIRUS, AND THEIR  
APPLICATIONS



This application is a continuation-in-part of PCT Application No. PCT/FR00/01742, filed June 21, 2000, which designated the U.S., the entire content of which is incorporated herein by reference.

5       The invention relates to the means, tools and methods for identifying the locus of a major resistance gene to the Rice Yellow Mottle Virus (RYMV) in short. It more particularly relates to markers and PCR primers in respect of tools.

10       RYMV is a virus that is endemic in Africa. In a few rare varieties of the African species of cultivated rice *Oryza glaberrima*, a very high resistance to RYMV has been identified. But since the interspecific hybrids between the two species of cultivated rice are  
15 extremely sterile, prior research has not been able to describe either the genetic bases or the mechanism of this resistance.

20       Research by the inventors in this area has shown that a variety called *Gigante* which originated from Mozambique and was identified by ADRAO, and which is a member of the cultivated Asian rice species *Oryza sativa*, shows the same characteristics as those observed with *O. glaberrima*. The inventors have characterized RYMV resistance by demonstrating that it

is related to a major recessive resistance gene that is identical in both sources of resistance under consideration (*O. Sativa* and *O. glaberrima*).

5 This resistance occurs at the level of cell-to-cell movement and leads to blockage of the virus at the infected cells whereas virus replication is normal.

On the contrary, in resistant varieties, a mutation of said protein does not enable anymore the association with the virus, and thus its diffusion in  
10 the plant.

The migration of RYMV occurs under the form of a nucleoproteic complex associating viral nucleic acid, of protein and movement virus protein. In the cells of sensitive varieties, an host factor, probably a  
15 protein, also contributes to the movement of the virus.

Having regard to these results, the inventors prepared specific method and tools for characterizing the chromosome fragment bearing said resistance to RYMV gene which codes for said protein enabling the movement  
20 of the virus in the plant.

The purpose of the invention is therefore to provide a method for identifying molecular markers of the resistance locus to RYMV.

It also concerns the DNA fragments such as  
25 revealed by this method and which can be used as markers.

The invention also concerns applications of such markers, in particular to define other markers having high specificity to the resistance locus and to predict  
30 a resistant phenotype.

The invention further relates to sequences of primers, as new products, used in the PCR techniques applied.

According to the invention, the identification of markers of the locus of a major resistance gene to RYMV, comprises the use of AFLP markers (Amplified Fragments Length Polymorphism) and uses the PCR  
5 technique.

This method of identification is characterized in that it comprises:

- selective amplification of rice DNA fragments firstly from resistant individuals and secondly from  
10 sensitive individuals, descending from parent varieties, these fragments being previously submitted to a digestion step, followed by ligation to fix complementary primer adapters having, at their end, one or more specific nucleotides, one of the primers in the  
15 primer pair being labelled for development purposes,
- separating the amplification products by gel electrophoresis under denaturing conditions, and
- comparing the electrophoresis profiles obtained with mixtures of fragments derived from resistant  
20 descendants and with mixtures derived from sensitive descendants, with the fragments derived from parent varieties, for the purpose of identifying bands whose polymorphism is genetically linked to the resistance locus, this identification being optionally followed,  
25 for validation purposes, by verification on each of the individuals and by calculation of the genetic recombination rate between the marker and the resistance locus.

In one embodiment of the invention, the DNA  
30 fragments are obtained by digestion of the genomic DNAs of resistant plants and of sensitive plants, and their parents, using restriction enzymes.

Restriction enzymes which have proved to be suitable include EcoRI and MseI.

Short nucleotide sequences are fixed to digestion fragments (adapters) to generate blunt ends to which  
5 the adapters are subsequently fixed.

The primers used in the amplification step are complementary to these adapters with, at their 3' end, from 1 to 3 nucleotides which may be variable.

The amplification step is advantageously conducted  
10 using the PCR technique.

Specific amplification profiles are obtained with primer pairs respectively having AAC and CAG, ACC and CAG motifs at their end, or further AGC and CAG.

The sequences corresponding to the EcoRI and MseI  
15 adapters are respectively GAC TGC GTA CCA ATT C (SEQ ID N°1) and GAT GAG TCC TGA GTA A (SEQ N°2).

The primer pairs used for amplification are then advantageously chosen from among E-AAC/M-CAG; E-ACC/M-CAG; and E-AGC/M-CAG; in which E and M respectively  
20 correspond to SEQ ID N°1 and SEQ ID N°2. Other pairs are given in table 6 in the examples.

Comparative study of the amplification profiles obtained reveals polymorphic bands specifically present in the sensitive varieties and their sensitive  
25 descendants, as shown in the examples, and consequently corresponding to resistance markers.

In particular, development by gel electrophoresis under denaturing conditions leads to identifying 2 marker bands M1 and M2 of respectively 510 bp and 140  
30 bp.

According to analysis of segregation data, these 2 bands determine a chromosome segment of 10 to 15 cM

carrying the resistance locus and are located either side of this locus at 5-10 cM.

According to one provision of the method of the invention, the polymorphic bands identified as markers  
5 specific to the RYMV resistance locus, are isolated from gels. Advantageously the electrophoresis gels are excised. This isolation step is followed by purification using conventional techniques. In this manner DNA fragments are obtained.

10 According to another provision of the invention, said purified fragments are cloned in an appropriate vector, such as a plasmid, inserted into the host cells, in particular bacterial cells such as those of *E.coli*.

15 According to another provision of the invention, the purified, cloned DNA fragments are sequenced.

Taking advantage of the sequences of the inserts corresponding to said DNA fragments, the invention also provides a method for obtaining markers having high  
20 specificity for the locus of a major resistance gene to RYMV. This method is characterized in that PCR primer pairs are determined which are complementary to the fragments of the sequence of a given insert, specific amplification of the insert is made using these primer  
25 pairs, and the amplification products are then subjected to migration on electrophoresis gel.

These DNA sequences can be used to identify a polymorphism linked to the resistance locus in a rice variety to be examined using different methods as  
30 described in the examples:

1) by directly identifying a size polymorphism of these DNA sequences after specific amplification and separation of the fragments on agarose gel,

2) by digesting the amplification products with restriction enzymes to separate the digestion products on agarose gel,

3) by using these sequences as probes to  
5 hybridize the DNA of rice varieties previously digested by a restriction enzyme and to determine a restriction polymorphism.

The invention concerns, as new products, the polymorphous AFLP bands such as identified by the  
10 method defined above, from the DNA of rice plants, optionally isolated, purified and sequenced.

These AFLP bands are characterized in that they are specifically revealed in a variety sensitive to RYMV (IR64) and in the fraction of sensitive plants  
15 derived from the crossing of this variety with the *Gigante* resistance variety as described in the examples.

The invention particularly concerns the DNA sequences corresponding to these polymorphous bands,  
20 which can be used to define a segment of chromosome 4 of 10-15 cM carrying the resistance locus to RYMV.

Having regard to the method with which they are obtained, the AFLP bands correspond to restriction fragments and in particular, according to one  
25 embodiment of the method of the invention, to EcoRI-MseI fragments.

Fragments of this type are called M1 and M2 markers and are characterized by a size, of 510 bp and 140 bp respectively, in electrophoresis gel under  
30 denaturing conditions.

These fragments are characterized in that they correspond to DNA sequences flanking the resistance locus and located either side of the latter at 5-10 cM.

The invention also concerns fragments cloned in vectors such as plasmids, these cloning vectors as such, characterized in that they comprise such fragments, and the host cells transformed using these  
5 vectors, such as bacterial cells, for example *E. coli*.

The invention relates in particular to the DNA sequence corresponding to the fragment identified as M1 marker and meeting the following sequence SEQ ID N°3:

10 CGTGCTTGCTTATAGCACTACAGGAGAAGGAAGGGGAACACAACAGC  
CATGGCGAGCGAAGGTTCAACGTCGGAGAAACAGGCTGCGACGGGCA  
GCAAGGTGCCGGCGGGCGGATCGGAGGAAGGAAAAGGAGGAAATCGA  
AGTTATGCTGGAGGGGCTTGACCTAAGGGCAGATGAGGAGGAGGATG  
TGGAATTGGAGGAAGATCTAGAGGAGCTTGAGGCAGATGCAAGATGG  
15 CTAGCCCTAGCAACAGTTCATACGAAGCGATCGTTTAGTCAAGGGGCT  
TTCTTTGGGAGTATGCGCTCAGCATGGAACTGCGCGAAAGAAGTAGAT  
TTCAGAGCAATGAAAGACAATCTGTTCTCGATCCAATTCAATTGTTTG  
GGGGATTGGGAACGAGTTATGAATGAAGGTCCATGGACCTTTCGAGG  
ATGTTTCGGTGCTCCTCGCAGAATATGATGGCTGGTCCAAGATTGAAT

20

The DNA sequence of the M1 marker has a size of 471 bp.

The invention also concerns, as new products, the sequences of nucleotides used as PCR amplification  
25 primers.

Such primers comprise the pairs E-AAC/M-CAG; E-ACC/M-CAG; E-ACC/M-CAG; in which E and M respectively relate to SEQ ID N°1 and SEQ ID N°2.

Other primers are complementary to sequences  
30 identified in the sequence of the fragment designated by marker M1. These are in particular (5',3') sequences chosen from among:

AGGAAGGGGAACACAACAGCC (21 bp) (SEQ ID N°4)



TTATGCTGGAGGGGCTTGACC (21 bp) (SEQ ID N°5)

GCAGTTCCATGCTGAGCGCAT (21 bp) (SEQ ID N°6)

CCGAACATCCTCGAAAGGTCC (21 bp) (SEQ ID N°6)

TCATATTCTGCGAGGAGCACC (21 bp) (SEQ ID N°8)

5       The invention also concerns the DNA sequence corresponding to the fragment identified as marker M2 and corresponding to sequence SEQ ID N°9

AATTCACCCC ATGCCCTAAG TTAGGACGTT CTCAGCTTAG TGGTGTGGTA  
GCTTTTTCTA TTTTCCTAAG CACCCATTGA AGTATTTTGC ATTGGAGGTG

10 GCCTTAGGTT TGCCTCTGTTA

      The size of M2 is 120 bp.

      Specific primers complementary to sequences identified in the sequence of M2 were defined. Said sequences meet the following sequencing (5',3'):

15 SEQ ID N°10

AACCTAAGGCCACCTCCAAT

SEQ ID N°11

GCAAACCTAAGGCCACCTC

SEQ ID N°12

20 ATTCACCCCATGCCCTAAG

      According to a further aspect of the invention, the latter concerns the use of DNA sequences obtained with the above primers to define polymorphisms which can be used to identify resistant phenotypes.

25       The invention also concerns a method for identifying the DNA sequence carrying the major resistance gene to RYMV. This method is characterized by screening a bank consisting of DNA fragments of 100 to 150 kb of the IR64 or other variety, such as the BAC  
30 bank (Bacterial Artificial Chromosomes) cloned in bacteria, to select the clone or clones from the bank containing the markers defined above and the resistance gene to RYMV.

This type of BAC bank is available from the IRRI institute.

The existence of different restriction sites on the sequence corresponding to the M1 marker, and in particular the sites corresponding to HpaII/MspI, provides for advantageous identification of resistant phenotypes.

The identification of different restriction sites on the sequence corresponding to the M1 marker enables characterization of a polymorphism which may be put to advantageous use to map the M1 marker on rice genetic linkage maps.

The map of the sequence corresponding to the M1 marker can be used to identify a chromosomal zone on chromosome 4 of rice carrying the RYMV resistance locus.

The map of the RYMV resistance gene on chromosome 4 of the rice genetic map allows identification of the markers the closest to the resistance locus. These are in particular the microsatellite markers RM252 and RM273 or any other marker inside the (4-5cM) space defined by these markers allowing identification of a polymorphism between the IR64 and Gigante parents, such as the RFLP markers screened from genomic banks or cDNA, microsatellites, AFLP markers or markers derived from physical mapping of the region such as BAC, YAC clones or their cosmids.

The markers identified in accordance with the invention, or any other marker located in this space allowing identification of a polymorphism between resistant varieties such as Gigante or O. Glaberrima with RYMV-sensitive rice varieties, may be used for transfer of RYMV resistance into sensitive varieties by

successive backcrosses followed by marker-assisted selection.

Other characteristics and advantages of the invention will be given in the following examples, in which reference is made to figures 1 to 10 which respectively represent:

- figure 1: cloning of marker M1 in the PGEMTeasy plasmid. Digestion of the plasmid shows a DNA fragment of 510 bp corresponding to band M1;

10 - figure 2: amplification of marker M1 in the four rice varieties (*Azucena*, *Gigante*, IR64 and Tog5681) using the primer pairs (2-4): 291 bp; (2-5): 310 bp; (1-3): 288 bp; (1-4): 406 bp; (1-5): 425 bp; (2-3). The M1 fragment is slightly bigger in Tog5681 than in the  
15 other varieties;

- figure 3: identification of restriction sites on the sequence of the M1 marker in the 4 varieties IR64, *Azucena*, *Gigante* and Tog5681;

- figure 4: digestion of the M1 marker with the  
20 HpaII enzyme after PCR amplification using primer pairs (1-3), (1-4) and (1-5) on the four varieties (*Azucena*, *Gigante*, IR64 and Tog5681). The presence of a HpaII restriction site in the IR64 and Tog568 varieties releases a fragment of 86 bp which reduces the size of  
25 the amplified fragment to the same extent.

- figure 5: characterization of the M1 marker on sensitive and resistant plants of F2 issue (IR64 and *Gigante*). The resistant F2 plants have the profile of the resistant parent (IR64 - no HpaII site), with the  
30 exception of a single recombinant, the resistant plants have the profile of the sensitive parent (IR64 - presence of HpaII site) with the exception of two recombinants;

- figure 6: segregation of the M1 marker in the HD population (IR64 x Azucena); IR64-Azucena-30 HD individuals (IR64 x Azucena);

5 4 - figure 7: the genetic linkage map of chromosome of rice with the positioning of marker M1 and identification of the space interval in which the resistance locus is found;

10 - figure 8: hybridization of M1 marker used as probe on membranes carrying the DNA of the 4 varieties (IR64, Azucena, Gigante and Tog5681) digested by 6 restriction enzymes ApaI, KpnI, PstI, Scal, HaeIII. The Tog5681 variety shows a different restriction profile to the other varieties for the Scal enzyme which may be used to label the resistance locus of this variety; and

15 - figure 9: hybridisation of the M1 marker used as probe on membranes carrying the DNA of individuals derived from backcross (IR64 x Tog568) x Tog 5681 and digested with the Scal enzyme. These descendants are in segregation for RYMV resistance. The sensitive  
20 individuals (5) all show the IR64 band associated with the Tog5681 band (heterozygote individuals). The resistant individuals (9) only show the Tog5681 band with the exception of one recombinant individual,

25 - figure 10: mapping and anchoring of the locus of bred resistance to RYMV on the map IR64 x Azucena, and

30 - figure 11, the genetic map of the region flanking the resistance gene in the IR64 x Gigante population (figure 11A) and the simplified representation of contig 89 and of part of the clones assigned to this contig.

Example 1: Identification of resistant-source varieties

The varieties used in the resistance study, and especially the two resistant varieties *Gigante* and Tog5681, were characterized using microsatellite markers on a representative sampling of loci.

- 5 Polymorphism is evidenced by the number of repeats of a short nucleotide pattern, most often binucleotide which is characteristic of a given variety.

On a set of loci, the catalogued alleles can provide specific characteristics for each variety.

- 10 The detection of these microsatellite markers is made by DNA amplification using the specific primers determined by Chen et al (1) followed by migration on polyacrylamide gel under denaturing conditions in accordance with the protocol described by the same  
15 authors.

- Table 1 gives the results using a reference system drawn up by Chen et al above, according to which the alleles are identified by the number of pattern repeats compared with the IR36 variety used as control. The two  
20 varieties *Gigante* and Tog5681 are therefore specifically described on 15 loci in respect of any other varieties (the microsatellite markers are given in column one).

Table 1

Locus	Chr	Size on IR36	Ref.	IR36	Gigante	IR64	Azucena	Tog5681
RM001	1	113	(2)	n	n-26	n	n-22	n-26
RM005	1	113	(2)	n	n-6	n-4	n+16	n-8
RM011	7	140	(2)	n	n-4	n	n-24	n-16
RM018	7	157	(2)	n	n+4	n+6	n+8	n-6
RM019	12	226	(2)	n	n	n+21	n-9	n-21
RM021	11	157	(2)	n	n+8	n	n-14	n-32
RM148	3	129	(3)	n	n+6	n	n	n+6

RM167	11	128	(3)	n	n+4	n	n+32	n+24
RM168	3	116	(3)	n	n-20	n	n-20	n-24
RM232	3	158	(1)	n	n-14	n	n-12	n-16
RM022	3	194	(2)	n	n-2	n	n-4	n-2
RM252	4	216	(1)	n	n+38	n+2	n-20	n+10
RM255	4	144	(1)	n	n	n	n	n
RM246	1	116	(1)	n	n-12	n-12	n-16	n-12
RM231	3	182	(1)	n	n+6	n-22	n-4	n-12

#### Example 2: Characterization of resistance

Resistance was characterized using artificial inoculation of young seedlings with the virus, compared  
5 with an extremely sensitive control variety IR64.

The virus content was followed up for 60 days after inoculation using ELISA tests on the most recent leaves.

10 These tests were never able to demonstrate a signal that was significantly different to the signal of control plants non-inoculated with the virus.

A further experiment was conducted by inoculating isolated protoplasts of the two varieties Tog5681 and Gigante. In both cases, it was possible to detect the  
15 presence of viral proteins (capsid protein and P1 movement protein) and the accumulation of viral DNA, demonstrating the capacity of these protoplasts to multiply the virus, in the same manner as the protoplasts of sensitive varieties such as IR64.

20 Therefore, if it is considered that replication, cell-to-cell movement and long-distance transport through the vessels are the three main steps in the process of the infectious cycle within the plant, the resistance of these two varieties most logically lies  
25 in blockage of the virus at the infected cells.

Example 3: Resistance genetics

Different F1 crosses were made between the resistant *O. sativa* variety (Gigante), a resistant *O.*  
5 *glaberrima* variety (Tog5681 - also identified by ADRAO), and the highly sensitive control variety IR64 (selected at the IRRI).

Culture of the plant material, crosses and production of descendants were made in the IRD  
10 greenhouses in Montpellier.

The F1 hybrids obtained between the sensitive and resistant varieties were tested for resistance to the RYMV virus by ELISA testing and follow-up of symptoms.

These F1 hybrids proved to be as sensitive as the  
15 sensitive parent, and therefore showed that the type of resistance is recessive.

On the other hand, the hybrids between the two resistance sources Gigante and Tog5681 only yielded resistant F1 hybrids to the benefit of a single  
20 resistance locus in these sources of resistance.

These results are summarized in Table 2 below.

This table gives the distribution of ELISA responses (A 405 nm) in the leaves infected by systemic route of F1 hybrids, of backcrosses and of F2  
25 descendants obtained from backcrosses between the sensitive IR64 variety and the 2 resistant cultivars Gigante and Tog5681.

TABLE 2

F1 hybrid descendants	Presence of symptoms	Number of genotypes	Distribution of OD values			Average values
			(0.01 - 0.05)	(0.9 - 1)	> 1	
Derivatives of Tog5681						
F1: (IR64 x Tog 5681)	Sensitive	-	-	-	10	1.9
BCS: (IR64 x Tog 5681) x IR64	Sensitive	19	6	4	15	1.6
BCS: (IR64 x Tog5681 c Tog5681	In segregation	22	12	-	10	-
Derivatives of fertile BCS plant						
BCS F2	Sensitive	11	-	-	11	1.3
BCS x IR64	Sensitive	1	-	-	1	1.9
BCS x Tog5681	sensitive	15	-	-	15	1.9
Gigante derivatives						
F1 (IR64 x Gigante)		-	-	-		
F2: (IR64 x Gigante)	In segregation	65	15	-	10	1.9
F1: (Gigante x Tog5681)	Sensitive	-	10	-	50	-
					-	0.3

The ELISA responses were obtained from:

- i) 10 plants regenerated by cuttings for each F1 hybrid combination
- ii) 1 plant regenerated for each backcross-derived interspecific genotype
- iii) direct tests on young seedlings (inoculation at 10 days after germination and read-off at 7 days after inoculation) for F2 and fertile interspecific descendants



In respect of *Gigante*, the heredity of resistance was confirmed by a resistance test on 55 F3 families resulting from the cross between (IR64 x *Gigante*). The results are given in Table 3.

This table gives the segregation of RYMV resistance in F3 descendants (IR64 x Gigante). Inoculation was made 10 to 17 days after germination with the Burkina Faso isolate and symptoms were followed up for 45 days after inoculation.

TABLE 3

Classes of resistance	Number of descendants	Number of plants			Incidence of resistant plants	
		Total	Sensitive	Resistant		
Sensitive	15	191	191	0	0	
In segregation	30	343	262	01	0.24	
Resistant	4	45	14	31	2 = 0.07 (3:1)	
Very resistant	6	87	0	87	0.69	
Resistant*	7	73	23	50	1	
Very resistant*	4	56	0	56	0.60	
					1	

\*F3 descendants derived from resistant F2 plants analysed by ELISA tests

Examination of this table shows that:

- ¼ of F2 plants only give resistant plants in F3 descendants, and are homozygote for resistance,
- ¼ of F2 plants only give sensitive plants in F3 descendants, and are homozygote for sensitivity,
- ½ of F2 plants are in segregation for resistance and give sensitive and resistant plants in the same proportion (3:1) in F3 descendants.

All these results tally perfectly with a single recessive resistance gene occurring in the two varieties *Gigante* and *Tog5681*.

Example 4: Identification M1 and M2 resistance markers using the AFLP protocol

15

a - Obtaining DNA pools

The leaves of 10 sensitive plants and 10 resistant plants derived from an F2 (*IR64* x *Gigante*) were sampled for their DNA extraction.

20 The DNA were then mixed stoechiometric fashion to form two DNA pools respectively corresponding to 10 sensitive or resistant F2 plants and with a final mixture concentration of 50 ng/µl. These mixtures served as basis for the identification of resistance markers using the AFLP (Amplified Fragments Length Polymorphism) method developed by Zaneau et al (4) and Vos et al (5). The products used are in the form of a commercial kit (Gibco BRL) available from Keygene & Life Technologies.

30 b - Obtaining restriction fragments

250 ng of each of the DNA pools at 50 ng/µl and of the parents are digested simultaneously by two restriction enzymes (*EcoRI* and *MseI*).

Digestion reaction (25  $\mu$ l):

- 5  $\mu$ l DNA (50 ng/ml)
- 0.2  $\mu$ l (2 U) EcoRI (10U/ $\mu$ l)
- 5 0.2  $\mu$ l (2 U) MseI (5U/ $\mu$ l)
- 5  $\mu$ l 5X T4 ligase buffer
- 14.5  $\mu$ l H<sub>2</sub>O

The digestion reaction is carried out for two hours at 37°C, then for 15 min at 70°C to inactivate the restriction enzymes. After digestion, the ligation reaction was performed.

Ligation reaction (50  $\mu$ l):

- 25  $\mu$ l double digestion reaction medium
- 15 1  $\mu$ l EcoRI adapter
- 1  $\mu$ l MseI adapter
- 5  $\mu$ l 5X T4 ligase buffer
- 1  $\mu$ l (1 U) ligase (10 U/ $\mu$ l)
- 17  $\mu$ l H<sub>2</sub>O

20 The ligation reaction is conducted at 37°C for 3 hours followed by inactivation of the enzyme at 60°C for 10 min.

c - Amplification

- 25 Amplification properly so-called was performed in two steps: preamplification and specific amplification.

c1 - Preamplification reaction (50 $\mu$ l)

- 30 5  $\mu$ l of reaction medium containing the digested DNA fixed to the adapters, diluted to 1/10
- 0.5  $\mu$ l EcoRI primer (150 ng/ $\mu$ l)

- 2  $\mu$ l 5mM nucleotide mixture  
5  $\mu$ l 10 X buffer, Promega  
5  $\mu$ l  $MgCl_2$ , 25 mM  
0.2  $\mu$ l (1 U) Taq polymerase (5 U/ $\mu$ l)  
5 31.8  $\mu$ l  $H_2O$

The characteristics of PCR pre-amplification are the following:

- 20 cycles with      denaturing: 30 sec at 94°C  
10                      hybridization: 30 sec at 56°C  
                         elongation: 1 min at 72°C

Selective amplification is made using an aliquot of the first amplification diluted to 1/30 using primers having 3 selective nucleotides at the 3' end, and by labelling one of the primers to develop bands on  
15 autoradiography film.

The following primer pairs are used:

E-AAC/M-CAG

E-ACC/M-CAG

E-AGC/M-CAG

20 in which

E meets the sequence:

GAC TGC GTA CCA ATT C (SEQ ID N° 1), and

M meets the sequence:

GAT GAG TCC TGA GTA A (SEQ ID N°2)

25 The hybridization temperature is reduced by 0.7°C per cycle, throughout the 11 following cycles:

- last 20 cycles:      denaturing: 30 sec at 90°C  
                         hybridization: 30 sec at 56°C  
                         elongation: 1 min at 72°C

30 The EcoRI primer is labelled (for 0.5  $\mu$ l tube):

0.18  $\mu$ l EcoRI primer (5ng)

0.1  $\mu$ l  $\gamma^{33}P$  ATP (10 mCu/ $\mu$ l)

0.05 µl 10 X kinase buffer

0.02 µl (0.2U) T4 polymerase kinase (10U/µl)

0.15 µl H<sub>2</sub>O

The labelling reaction is conducted at 37°C for 1  
5 hour and is halted by 10 minutes at 70°C

c2 - Specific amplification reaction

(20 µl):

0.5 µl labelled EcoRI primer

10 5 µl preamplification reaction medium, diluted to  
1/30

0.3 µl MseI primer (100ng/µl)

0.8 µl 5mM nucleotide mixture

2 µl 10 X buffer, Promega

15 2 µl MgCl<sub>2</sub>, 25 mM

0.1 µl (0.5 U) Taq polymerase (5 U/µl)

9.3 µl H<sub>2</sub>O

Amplification characteristics are as follows:

32 cycles with

20 - for the first cycle:

denaturing: 30 sec at 94°C

hybridization: 30 sec at 65°C

elongation: 1 min at 72°C

- for the 11 following cycles: the same conditions as  
25 previously, reducing the hybridization temperature by  
0.7°C for each cycle; and

- for the 20 last cycles:

denaturing: 30 sec at 90°C

hybridization: 30 sec at 56°C

30 elongation: 1 min at 72°C

d) Electrophoresis and Autoradiography

At the end of the amplification reaction, 20 µl of charge buffer are added (98% formamide, 0.005 % xylene cyanol and 0.005 % bromophenol blue). The amplification products are separated by electrophoresis on denaturing polyacrylamide gel (6% acrylamide, 8 M urea) with a TBE migration buffer (18 mM Tris, 0.4 mM EDTA, 18 mM boric acid, pH 8.0) for 3 hours' migration at a power of 50 watts. After migration, the gel is fixed in a solution of 1 part acetic acid/ 2 parts absolute ethanol for 20 minutes. The gel is transferred to 3M Wattman paper and dried for 45 minutes at 80°C with a gel drier. The gel is placed in a cassette with ultrasensitive film. The autoradiograph is developed after two days' exposure. Comparison of the profiles obtained with the parents and the pools of sensitive of resistant plants led to identifying bands present in one of the pools but absent in the other. These bands, candidates for resistance marking, were then verified individually on each of the plants forming the DNA pools.

e) Results

Study of the results obtained shows that the two markers called M1 and M2 are present in the sensitive parent (IR64) and in all F2 plants (IR64 x *Gigante*) forming the pool of sensitive plants, whereas this band is absent in the resistant parent (*Gigante*) and that only one individual in the resistant pool shows this band. The same type of variation is observed in backcross (IR64 x Tog55681) x Tog 5681. The other markers identified by this analysis (M3 to M6) also show the same variation:

- presence of bands in the sensitive parent and the pool of F2 sensitive plants (IR64 x *Gigante*) and in the sensitive plants of the backcross (IR64 x Tog5681) x Tog5681).

- 5        - absence of bands in the resistant parents *Gigante* and Tog5681, in the pool of F2 resistant plants (IR64 x *Gigante*) and in the resistant plants of the backcross (IR64 x Tog5681) x Tog5681.

The segregation data between the AFLP markers M1  
10 to M6, the resistance locus for the F2 pools (IR64 x *Gigante*) and the interspecific backcross (IR64 x Tog5681) x Tog5681 are summarized in tables 4 and 5. Analysis of the segregation data and of the rare recombinants observed in both crosses can be used to  
15 assess the recombination rates between these different markers and the resistance locus. In particular, markers M1 firstly and markers M2 to M6 secondly determine a segment of less than 10-15 cM carrying the resistance locus. M1 and M2 are therefore less than 5-  
20 10 cM apart and are positioned either side of this locus.

TABLE 4

Resistance/Marker M1	N° of individuals observed						
Phenotype	Resistant			Sensitive			
RYMV resistance genotype	<i>tt/gg</i>	<i>tt</i>	<i>gg</i>	<i>It</i>	<i>It</i>	<i>It</i>	<i>It</i>
AFLP marker	-/-	+/-	+/	-/-	+/-	-/-	+/
Resistant F2 pool (IR64 x <i>Gigante</i> )	10	-	1	-	-	-	-
Sensitive F2 pool (IR64 x <i>Gigante</i> )	-	-	-	-	-	0	10
Interspecific backcross Tog5681	11	1	-	0	8	-	-
Resistance/Marker M2, M3, M4, M6	N° of individuals observed						
Phenotype	Resistant			Sensitive			



RYMV resistance genotype	<i>tt/gg</i>	<i>tt</i>	<i>gg</i>	<i>It</i>	<i>It</i>	<i>II</i>	<i>II</i>
AFLP marker	-/-	+/-	+/	-/-	+/-	-/-	+/
Resistant F2 pool (IR64 x <i>Gigante</i> )	11	-	0	-	-	-	-
Sensitive F2 pool (IR64 x <i>Gigante</i> )	-	-	-	-	-	0	10
Interspecific backcross Tog5681	10	2	-	0	8	-	-
Resistance/Marker M5	N° of individuals observed						
Phenotype	Resistant			Sensitive			
RYMV resistance genotype	<i>tt/gg</i>	<i>tt</i>	<i>gg</i>	<i>It</i>	<i>It</i>	<i>II</i>	<i>II</i>
AFLP marker	-/	+/-	+/	-/-	+/-	-/-	+/
Resistant F2 pool (IR64 x <i>Gigante</i> )	11	-	-	-	-	-	0
Sensitive F2 pool (IR64 x <i>Gigante</i> )	-	-	-	-	-	0	10
Interspecific backcross Tog5681	9	3	0	8	-	-	-

TABLE 5

Marker M1/Markers M2,M3,M4,M6	N° individuals observed			
Genotype M1	-/*	+/*	-/-	-/-
Genotype M2,M3,M4,M6	+/*	-/-	+/*	-/-
Resistant F2 pool (IR64 x <i>Gigante</i> )	0	1	0	10
Sensitive F2 pool (IR64 x <i>Gigante</i> )	10	0	0	0
Interspecific backcross Tog5681	11	2	2	11
Marker M1/Marker M5	N° individuals observed			
Genotype M1	-/*	+/*	-/-	-/-
Genotype M5	+/*	-/-	+/*	-/-
Resistant F2 pool (IR64 x <i>Gigante</i> )	0	1	0	10
Sensitive F2 pool (IR64 x <i>Gigante</i> )	10	0	0	0
Interspecific backcross Tog5681	11	2	3	10
Marker M5/Markers M2,M3,M4,M6	N° individuals observed			

Genotype M5	+/*	+/*	-/-	-/-
Genotype M2,M3,M4,M6	+/*	-/-	+/*	-/-
Resistant F2 pool (IR64 x <i>Gigante</i> )	0	0	0	11
Sensitive F2 pool (IR64 x <i>Gigante</i> )	10	0	0	0
Interspecific backcross Tog5681	13	1	0	12

\*: (-) interspecific backcross Tog5681 (+ or -) F2 pool.

#### Example 5: Isolation of marker M1

5        A further amplification with the same pair of primers was conducted, followed by migration on polyacrylamide gel under the same conditions as above. Development was carried out by staining with silver nitrate using the silver staining kit (Promega) for  
10       direct viewing of the bands on the gel. After development, the M1 band was excised from the gel, then the DNA was eluted in 50 µl water at 4°C overnight.

15       An aliquot of 5 µl was taken and re-amplified using the same primer pairs with P<sup>33</sup> labelling. The amplification product was again separated on 6% denaturing acrylamide gel and compared with the parents and the sensitive and resistant pools. The lane corresponding to this amplification product shows a single band of 510 bp migrating at exactly the same  
20       level as the original band which had been excised. Another 5 µl aliquot was also amplified with the same primers and separated on 1.8% agarose gel. The band corresponding to the expected size (510 bp) was again excised and purified with a gene clean kit (Promega).

Example 6: Cloning and Sequencing of the M1 Marker- cloning

3 µl of purification product was used for a cloning reaction overnight at 37°C

- 5        3 µl purification product
- 1 µl PGEMTeasy vector
- 1 µl 10 X T4 ligase buffer
- 1 µl T4 DNA Ligase
- 4 µl H<sub>2</sub>O

- 10       Transformation was conducted with the *E. Coli* strain JM109, adding 5 µl of cloning product to 100 µl competent *E. Coli* JM109 cells. A pre-culture was made on LB culture medium for 1 hour at 37°C. The bacteria were subsequently spread over a Petri dish containing
- 15       agar with 1/1000 ampicilline. 50 µl IPTG-XGal were added just before spreading the bacteria to select the transformed bacteria. A white colony ( transformed) was selected and replaced in culture under the same conditions (Agar plus ampicilline).

- 20       From this culture a miniprep of plasmid DNA was MADE using the Wizard Plus kit (Promega). The plasmid DNA containing the insert was digested with the EcoRI enzyme to verify the presence of the M1 marker. 1.8% agarose gel was used to verify the presence of the 3 kb
- 25       band corresponding to the plasmid and the 510 bp band corresponding to the M1 marker (photo 1).

- Sequencing

- The sequence of the insert (SEQ ID N°3) is the following (5',3'):
- 30

SED ID N°3

20       30       40       50       60       70

GTGCTTGCTTATAGCACTACAGGAGAAGGAAGGGGAACACAACAGCC  
 ATGGCGAGCGAAGGTTCAACGTCGGAGAAACAGGCTGCGACGGGCAG  
 CAAGGTGCCGGCGCGGATCGGAGGAAGGAAAAGGAGGAAATCGAA  
GTTATGCTGGAGGGGCTTGACCTAAGGGCAGATGAGGAGGAGGATGT  
 5 GGAATTGGAGGAAGATCTAGAGGAGCTTGAGGCAGATGCAAGATGGC  
 TAGCCCTAGCCACAGTTCATACGAAGCGATCGTTTAGTCAAGGGGCTT  
 TCTTTGGGAGTATGCGCTCAGCATGGAAGTGGCGGAAAGAAGTAGATT  
 TCAGAGCAATGAAAGACAATCTGTTCTCGATCCAATTCAATTGTTTGG  
 GGGATTGGAACGAGTTATGAATGAAGGTCCATGGACCTTTTCGAGGAT  
 10 GTTTCGGTGCTCCTCGCAGAATATGATGGCTGGTCCAAGATTGAAT

The sequences corresponding to the primers used  
 for AFLP amplifications were found and show that the  
 band corresponds to a restriction fragment (EcoRI-  
 15 MseI).

By deducing the sequences corresponding to the  
 primers, the actual size of the DNA fragment of the  
 cloned rice is 471 bp.

The use of different pairs of primers (1-3), (1-  
 20 4), (1-5) firstly and (2-3), (2-4), (2-5) secondly,  
 makes it possible to validate the cloning of the AFLP  
 M1 band. Amplification of the DNA of the varieties used  
 in the crosses with these primers only shows one single  
 band. The fragment corresponding to the Tog5681 variety  
 25 is slightly larger than for the other varieties  
 (fig.2).

#### Example 7: Transformation of the M1 sequence into a polymorphous marker

30

A polymorphism for the M1 marker was determined  
 between the parents of the doubled haploid population  
 (IR64 x Azucena). This population totals over 300

markers distributed over the 12 rice chromosomes. On this account, we relied on the restriction sites of the M1 marker sequence determined on the IR64 parent (fig.3). The primers (1-3), (1-4) and (1-5) were used to amplify the DNA of the parents of crossed plants which was then digested by restriction enzymes. The restriction site HpaII/MspI releases a fragment of 86 bp when primer 1 is used. This site is absent in the *Gigante* and *Azucena* varieties (fig. 4).

10       The marker was tested on the F2 individuals of the sensitive pool and resistant crossed pool (IR64 x *Gigante*). All the resistant individuals have the profile of the *Gigante* variety (absence of the M1 AFLP marker associated with absence of the restriction site HpaII/MspI) with the exception of individual (5.11).  
15       The sensitive individuals show the HpaII/MspI restriction site in the homozygote state like the IR64 variety with the exception of two heterozygote individuals which are recombinant (fig.5).

20       The sequence of the M1 marker which can be amplified with specific primers indeed corresponds to the M1 AFLP marker. Digestion by the HpaII/MspI enzyme leads to distinguishing between the allele derived from the sensitive parent (IR64) and from the resistant  
25       parent (*Gigante*).

      With these new data, it is possible to give back-up to the positioning of the resistance locus between markers M1 and M2 and to estimate the recombination rate at  $0.065 \pm 0.045$  for the distance between M1 and  
30       the resistance locus, and  $0.11 \pm 0.047$  for the distance between markers M1 and M2.

#### Example 8: Mapping of the M1 marker

Sixty individuals from the (IR64 x Azucena) population were passed as marker M1: amplification with primers (1-3) and digestion with the HpaII/MspI enzyme, followed by separation of the fragments on 2.5 % agarose gel. Segregation of marker M1 shows no distortion (fig.6). The results are used to map the M1 marker using mapping software (Mapmaker V3) which leads to positioning the M1 marker on chromosome 4 between the markers RG 163 and RG 214 (fig.7). This space represents the zone in which the RYMV resistance locus is located.

Example 9: Marking the resistance locus of the Tog5681 variety

The presence of the restriction site HpaII/MspI in the Tog5681 variety means that it is not possible to use the strategy in example 8 to verify that the M1 marker is also a marker of Tog5681 resistance derived from Tog5681. Therefore, the 4 varieties Azucena, Gigante, IR64 and Tog5681 were digested with 12 restriction enzymes (BamHI, BglII, DraI, EcoRI, EcoRV, HindIII, ApaI, KpnI, PstI, Scal, XbaI, HaeIII) to identify a restriction polymorphism using the DNA sequence of the M1 marker as probe. The Scal enzyme leads to identifying a polymorphism between IR64 and Tog5681 (fig.8). This polymorphism was used to validate the M1 marker on a backcross (IR64 x Tog5681) x IR64 in segregation for resistance. 5 sensitive individuals of this backcross were tested and all showed the characteristic band of IR64. The 9 resistant individuals only show the Tog5681 band with the exception of only one which is recombinant (fig.9). The restriction polymorphism revealed by the Scal enzyme

using the M1 marker as probe is therefore related to the resistance locus of Tog5681. There is coherence between genetic analysis and the identification of resistance markers for considering that the M1 marker  
 5 indeed maps the same resistance locus in the two varieties *Gigante* and Tog5681.

Example 10: Cloning and sequencing of the M2 marker into a specific PCR-marker.

10 The AFLP band obtained with the pair of primers E-ACC/M-CAG corresponding to the M2 band visible in the sensitive parent (IR64) and present in all the individuals forming the sensitive pool, was cloned using the same protocol as for marker M1. The sequence  
 15 corresponding to this band was determined and 3 primers were defined (1 forward - 2 reverse) to allow conversion of this marker into a specific PCR marker.

Sequence of the M2 marker (120 bp) (SEQ ID N°9):

AATTCACCCC ATGCCCTAAG TTAGGACGTT CTCAGCTTAG  
 20 TGGTGTGGTA GCTTTTCTA TTTTCCTAAG CACCCATTGA  
 AGTATTTTGC ATTGGAGGTG GCCTTAGGTT TGCCTCTGTTA

Primers:

(SEQ ID N°10): AACCTAAGGCCACCTCCAAT (right)  
 25 (SEQ ID N°11): GCAAACCTAAGGCCACCTC (right)  
 (SEQ ID N°12): ATTCACCCCATGCCCTAAG (left)

The following conditions were used to amplify markers M1 and M2 simultaneously:

30 - 10 X buffer, Promega 1.5 µl  
 - MgCl<sub>2</sub> Promega 1.5 µl  
 - dNTP (5 mM) 0.6 µl

	- M1-1 primer (10 mM)	0.15 µl
	- M1-4 primer (10 mM)	0.15 µl
	- M2-1 primer (10 mM)	0.15 µl
	- M2-2 primer (10 mM)	0.15 µl
5	- H <sub>2</sub> O	7.74 µl
	- Taq Polymerase	0.06 µl
	- DNA (5 ng/µl)	3.00 µl

PCR programme:

	- 5 min at 94°C
10	- 1 mn at 94°C
	- 30 s at 59°C
	- 1 mn at 72°C
	- 35 cycles
	- 5 mn at 72°C
15	- 10 mn at 4°C

The M2 marker may be amplified alone at a hybridization temperature of 60.5°C, the other parameters remaining unchanged. Under these amplification conditions, the M2 marker appears to be a dominant marker characterized by band presence in the sensitive parent (IR64) and band absence in the *Gigante* parent.

25 Example 11: Creation of a population of recombinant resistant plants between markers M1 and M2 to arrange within this space the candidate AFLP markers for resistance marking.

30 750 F2 individuals (IR64 x *Gigante*) were artificially inoculated with the RYMV virus (BF1 strain). The symptom-free plants were transplanted to a greenhouse, i.e. 188 individuals. Subsequently,



additional analysis based on ELISA and descendant tests made it possible to eliminate a last fraction of 50 sensitive plants. The remaining 138 plants, homozygote for resistance, were systematically genotyped for both markers M1 and M2 as previously described. In this manner, 45 individuals were selected (38 recombinant relative to M1. 7 recombinant relative to M2) and 2 double recombinants. These recombinant individuals were used for arranging the AFLP markers in the space between M1 and M2. These results are summarized in Table 6 below:

TABLE 6

Selection of a recombinant F2 sub-population (IR64 x Gigante) in the M1-M2 marker space

Steps conducted: F2 (IR64 x Gigante)	N° of plants	%
Inoculation of F2 plants(10 days after sowing)	768	
Greenhouse transplantation (5 weeks after inoculation)	188	
Elimination of sensitive plants (symptom follow-up - Elisa test, descendant test)	50	
Selection of homozygote resistant plants for the bred resistance gene	138	17.9
Genotyping of selected individuals for markers M1 and M2		
Recombinant plants relative to M1	36	18.8
Recombinant plants relative to M1 and M2	2	1.4
Recombinant plants relative to M2	7	5.1

Example 12: Screening of AFLP markers to select new candidate markers for resistance

A total of 328 primer pairs EcoRI/MseI, each one defined by 3 nucleotides, was used following the protocol previously described. These primers are given in Table 7 below.

TABLE 7

Combination	EcoRI	MseI	Combination	EcoRI	MseI	Combination	EcoRI	MseI
-------------	-------	------	-------------	-------	------	-------------	-------	------

Nº	primer	primer	Nº	primer	primer	Nº		primer
1	AAC	CAA	55	ACA	CTG	109	ACG	AGG
2	AAC	CAC	56	ACA	CTT	110	ACG	AGT
3*	AAC	CAG	57	ACA	AAC	111	ACT	CAA
4	AAC	CAT	58	ACA	AAG	112	ACT	CAC
5	AAC	CCA	59	ACA	AAT	113	ACT	CAG
6	AAC	CCT	60	ACA	ACA	114	ACT	CAT
7	AAC	CGA	61	ACA	ACC	115	ACT	CCA
8	AAC	CGT	62	ACA	ACG	116	ACT	CGT
9	AAC	CTA	63	ACA	ACT	117	ACT	CGA
10	AAC	CTC	64	ACA	AGC	118	ACT	CGT
11	AAC	CTG	65	ACA	AGG	119	ACT	CTA
12	AAC	CTT	66	ACA	AGT	120	ACT	CTC
13	AAC	AAC	67	ACC	CAA	121	ACT	CTG
14	AAC	AAG	68	ACC	CAC	122	ACT	CTT
15	AAC	AAT	69*	ACC	CAG	123	ACT	AAC
16	AAC	ACA	70	ACC	CAT	124	ACT	AAG
17	AAC	ACC	71	ACC	CCA	125	ACT	AAT
18	AAC	ACG	72	ACC	CCT	126	ACT	ACA
19	AAC	ACT	73	ACC	CGA	127	ACT	ACC
20	AAC	AGC	74	ACC	CGT	128	ACT	ACG
21	AAC	AGG	75	ACC	CTA	129	ACT	ACT
22	AAC	AGT	76	ACC	CTC	130	ACT	AGC
23	AAG	CAA	77**	ACC	CTG	131	ACT	AGG
24	AAG	CAC	78	ACC	CTT	132	ACT	AGT
25	AAG	CAG	79	ACC	AAC	133	AGA	CAA
26	AAG	CAT	80	ACC	AAG	134	AGA	CAC
27	AAG	CCA	81**	ACC	AAT	135	AGA	CAG
28	AAG	CCT	82	ACC	ACA	136	AGA	CAT
29	AAG	CGA	83	ACC	ACC	137	AGA	CCA
30	AAG	CGT	84	ACC	ACG	138	AGA	CCT
31	AAG	CTA	85	ACC	ACT	139	AGA	CGA
32	AAG	CTC	86**	ACC	AGC	140	AGA	CGT
33	AAG	CTG	87	ACC	AGG	141	AGA	CTA
34	AAG	CTT	88	ACC	AGT	142	AGA	CTC
35	AAG	AAC	89	ACG	CAA	143	AGA	CTG
36	AAG	AAG	90	ACG	CAC	144	AGA	CTT
37	AAG	AAT	91**	ACG	CAG	145	AGA	AAC

38	AAG	ACA	92	ACG	CAT	146	AGA	AAG
39	AAG	ACC	93	ACG	CCA	147	AGA	AAT
40	AAG	ACG	94	ACG	CCT	148	AGA	ACA
41	AAG	ACT	95	ACG	CGA	149	AGA	ACC
42	AAG	AGC	96	ACG	CGT	150	AGA	ACG
43	AAG	AGG	97	ACG	CTA	151	AGA	ACT
44	AAG	AGT	98	ACG	CTC	152	AGA	AGC
45	ACA	CAA	99	ACG	CTG	153	AGA	AGG
46	ACA	CAC	100	ACG	CTT	154***	AGA	AGT
47	ACA	CAG	101	ACG	AAC	155	AGC	CAA
48	ACA	CAT	102	ACG	AAG	156	AGC	CAC
49	ACA	CCA	103	ACG	AAT	157***	AGC	CAG
50	ACA	CCT	104*	ACG	ACA	158	AGC	CAT
51	ACA	CGA	105	ACG	ACC	159	AGC	CCA
52	ACA	CGT	106	ACG	ACG	160	AGC	CCT
53	ACA	CTA	107	ACG	ACT	161	AGC	CGA
54	ACA	CTC	108	ACG	AGC	162	AGC	CGT

Shaded: polymorphism for one or more bands between the sensitive and resistant pools

\* presence of one or more polymorphous bands in sensitive pool

5 \*\* presence of one or more polymorphous bands in resistant pool

\*\*\* presence of one or more polymorphous bands in sensitive pool and resistant pool

10

TABLE 7 (cont.)

Combination N°	EcoRI primer	MseI primer	Combination N°	EcoRI primer	MseI primer	Combination N°	EcoRI	MseI primer
163	AGC	CTA	218	AGT	AGC	273	CAT	CTA
164	AGC	CTC	219	AGT	AGG	274	CAT	CTC
165	AGC	CTG	220*	AGT	AGT	275	CAT	CTG
166	AGC	CTT	221	ATC	CAA	276	CAT	CTT
167	AGC	AAC	222	ATC	CAC	277	CAT	AAC
168	AGC	AAG	223	ATC	CAG	278	CAT	AAG
169	AGC	AAT	224	ATC	CAT	279	CAT	AAT
170	AGC	ACA	225	ATC	CCA	280*	CAT	ACA

171	AGC	ACC	226	ATC	CCT	281	CAT	ACC
172	AGC	ACG	227	ATC	CGA	282	CAT	ACG
173	AGC	ACT	228	ATC	CGT	283	CAT	ACT
174**	AGC	AGC	229	ATC	CTA	284	CAT	AGC
175***	AGC	AGG	230	ATC	CTC	285	CAT	AGG
176	AGC	AGT	231	ATC	CTG	286	CAT	AGT
177	AGC	CAA	232	ATC	CTT	287*	ACT	CAA
178	AAC	CAC	233***	ATC	AAC	288	CTA	CAC
179	AGG	CAG	234***	ATC	AAG	289	CTA	CAG
180	AGG	CAT	235*	ATC	AAT	290	CTA	CAT
181	AGG	CCA	236	ATC	ACA	291*	CTA	CCA
182	AGG	CCT	237	ATC	ACC	292	CTA	CCT
183	AGG	CGA	238	ATC	ACG	293	CTA	CGA
184	AGG	CGT	239	ATC	ACT	294	CTA	CGT
185	AGG	CTA	240	ATC	AGC	295	CTA	CTA
186	AGG	CTC	241	ATC	AGG	296	CTA	CTC
187	AGG	CTG	242	ATC	AGT	297*	CTA	CTG
188	AGG	CTT	243	CAA	CAA	298	CTA	CTT
189	AGG	AAC	244	CAA	CAC	299	CTA	AAC
190	AGG	AAG	245	CAA	CAG	300	CTA	AAG
191	AGG	AAT	246	CAA	CAT	301	CTA	AAT
192	AGG	ACA	247	CAA	CCA	302	CTA	ACA
193	AGG	ACC	248	CAA	CCT	303	CTA	ACC
194	AGG	ACG	249	CAA	CGA	304	CTA	ACG
195**	AGG	ACT	250**	CAA	CGT	305	CTA	ACT
196	AGG	AGC	251	CAA	CTA	306	CTA	AGC
197***	AGG	AGG	252	CAA	CTC	307	CTA	AGG
198	AGG	AGT	253	CAA	CTG	308	CTA	AGT
199	AGT	CAA	254*	CAA	CTT	309	CTT	CAA
200	AGT	CAC	255	CAA	AAC	310	CTT	CAC
201	AGT	CAG	256	CAA	AAG	311	CTT	CAG
202	AGT	CAT	257*	CAA	AAT	312**	CTT	CAT
203	AGT	CCA	258**	CAA	ACA	313	CTT	CCA
204	AGT	CCT	259	CAA	ACC	314	CTT	CCT
205	AGT	CGA	260	CAA	ACG	315	CTT	CGA
206	AGT	CGT	261	CAA	ACT	316	CTT	CGT
207	AGT	CTA	262	CAA	AGC	317	CTT	CTA
208	AGT	CTC	263	CAA	AGG	318*	CTT	CTC

209	AGT	CTG	264	CAA	AGT	319**	CTT	CTG
210	AGT	CTT	265	CAT	CAA	320	CTT	CTT
211	AGT	AAC	266	CAT	CAC	321	CTT	AAC
212	AGT	AAG	267	CAT	CAG	322	CTT	AAG
213*	AGT	AAT	268	CAT	CAT	323	CTT	AAT
214	AGT	ACA	269	CAT	CCA	324	CTT	ACA
215**	AGT	ACC	270	CAT	CCT	325	CTT	ACC
216	AGT	ACG	271	CAT	CGA	326	CTT	ACG
217	AGT	ACT	272*	CAT	CGT	327	CTT	ACT
						328	CTT	AGT

Shaded: polymorphism for one or more bands between the sensitive and resistant pools

\* presence of one or more polymorphous bands in sensitive pool

5 \*\* presence of one or more polymorphous bands in resistant pool

\*\*\* presence of one or more polymorphous bands in sensitive pool and resistant pool

With this screening, it was possible to identify  
 10 one or more polymorphous bands according to their occurrence in the sensitive parent and/or resistant parent. 23 primer pairs were able to identify polymorphism between the parents confirmed by the F2 DNA pools, sensitive or resistant. The table below  
 15 summarizes and gives the position in the M1-M2 space of the AFLP markers bound to the locus of bred resistance to the rice yellow mottle virus.

TABLE 8

Combination Number	Variable nucleotides		Presence of band(s)		Marker position in M1-M2 space
	EcoRI primer	MseI primer	Sensitive pool	Resistant pool	
3	AAC	CAG	-	-	=cloned M1 marker
69	ACC	CAG	+	-	=cloned M2 marker
77	ACC	CTG	-	+	non-determined
81	ACC	AAT	-	+	non-determined
86	ACC	AGC	-	+	non-determined

91	ACG	CAG	-	+	non-determined
104	ACG	ACA	+	-	betw. R and Rm273
154	AGA	AGT	+	+	beyond M2
157	AGC	CAG	-	+	in cosegr with M2
174	AGC	AGC	-	+	non-determined
175	AGC	AGG	+	+	betw M1 and Rm241
197	AGG	AGG	+	+	betw M1 and Rm241
215	AGT	ACC	-	+	non-determined
220	AGT	AGT	+	-	betw Rm273 and M2
233	ATC	AAG	+	+	betw M1 and Rm241
250	CAA	CGT	-	+	non-determined
254	CAA	CTT	+	-	beyond M2
258	CAA	ACA	+	-	betw M1 and Rm241
280	CAT	ACA	+	-	beyond M2
287	CTA	CAA	+	-	betw Rm273 and M2
291	CTA	CCA	+	-	betw M1 and Rm241
318	CTT	CTC	+	+	betw Rm273 and M2
319	CTT	CTG	-	+	non-determined

After separate verification on each of the individuals forming the pools, the candidate markers corresponding to bands present in the IR64 parent may be tested on the recombinants identified in example 11. In this manner, 9 markers were confirmed as belonging to the M1-M2 space. Table 9 gives the order in the M1-M2 space of the AFLP markers identified by comparing sensitive and resistant DNA pools from a resistant F2 sub-population (IR64 x Gigante).

TABLE 9

F2 Resistant individuals (IR64 x Gigante)	M1	E- AGG M- AGG	E- ATC M- AGG	E- CAA M- ACA	E- AGC M- AGG	E- CTA M- CCA	RM241	RM252	RYMV resist	E- ACG M- ACA	RM273	E- AGT M- AGT	C- CTT M- CTC	E- CTA M- CAA	M2
2	H	D	D	D	D	D	-	B	B	B	B	B	B	B	B
7	H	D	D	D	D	D	-	B	B	B	B	B	B	B	B

8	H	D	D	D	D	D	-	B	B	B	B	B	B	B	B
10	H	D	D	D	E	D	-	B	B	B	B	B	B	B	B
21	H	D	D	D	D	B	C	B	B	B	B	B	B	B	B
23	H	D	D	D	E	D	-	B	B	B	B	B	B	B	B
25	H	D	D	D	D	D	E	H	B	B	B	B	B	B	
28	H	D	D	D	B	B	C	B	B	B	B	B	B	B	B
37	H	D	D	D	E	D	E	H	B	B	B	B	B	B	B
48	H	D	D	D	D	D	-	B	B	B	B	B	B	B	B
55	H	D	D	D	D	D	E	H	B	B	B	B	B	B	B
61	H	D	D	D	D	D	E	H	B	B	B	B	B	B	B
65	H	D	D	D	-	B	C	B	B	B	B	B	B	B	B
95	H	E	D	D	D	B	C	B	B	B	B	B	B	B	B
103	H	E	D	D	-	B	C	B	B	B	B	B	B	B	B
104	H	D	D	D	B	B	C	B	B	B	B	B	B	B	B
109	H	B	B	B	B	B	C	B	B	B	B	B	B	B	B
111	H	E	D	D	D	D	-	B	B	B	B	B	B	B	B
119	H	D	D	D	D	D	-	B	B	B	B	B	B	B	B
120	A	D	D	D	D	B	C	B	B	B	B	B	B	B	B
125	H	E	E	E	E	D	-	B	B	B	B	B	B	B	B
127	H	-	-	-	-	B	C	B	B	B	B	B	B	B	B
131	H	E	E	E	E	D	-	B	B	B	B	B	B	B	B
133	H	-	-	-	-	B	C	B	B	B	B	B	B	B	B
141	H	E	E	E	E	D	E	H	B	B	B	B	B	B	B
154	H	E	E	E	E	D	-	B	B	B	B	B	B	B	B
158	H	E	E	E	E	D	-	B	B	B	B	B	B	B	B
159	H	-	-	-	-	B	C	B	B	B	B	B	B	B	B
160	H	E	E	E	E	D	-	B	B	B	B	B	B	B	B
151	H	E	E	E	E	D	-	B	B	B	B	B	B	B	B
153	H	-	-	-	-	B	C	B	B	B	B	B	B	B	B
157	H	-	-	-	-	B	B	B	B	B	B	B	B	B	B
171	H	-	-	-	-	B	B	B	B	B	B	B	B	B	B
175	H	E	E	E	D	D	B	B	B	B	B	B	B	B	B
179	H	-	-	-	B	B	B	B	B	B	B	B	B	B	B
183	H	E	E	E	E	D	B	B	B	B	B	B	B	B	B
35	H	D	D	D	D	D	H	H	B	D	H	D	D	D	D
135	H	E	E	E	E	D	H	H	B	B	H	D	D	D	D
17	H	B	B	B	B	B	-	B	B	D	H	D	D	D	D
20	B	B	B	B	B	B	B	B	B	D	H	D	D	D	D
38	B	B	B	B	B	B	-	B	B	D	H	D	D	D	D
93	B	B	B	B	B	B	B	B	B	D	H	D	D	D	D
105	B	B	B	B	B	B	B	B	B	D	H	D	D	D	D
145	B	-	-	-	B	B	B	B	B	B	B	B	B	B	D
180	B	-	-	B	B	B	B	B	B	B	B	D	D	D	D

Incidence of recombinant individuals \*

M1-R space    0.97   0.97   0.97   0.87 0.61   0.29  
 0.13  
 R-M2 space    0.67   0.78   0.89   0.89   0.89  
 Distance/resistance (cM) 11.4\*\*   11.03   11.03  
 5   11.03       9.88   6.90   3.33   2.10   0.00   3.33   3.89  
      4.44 4.44   4.44   5.0\*\*

- A:    genotype homozygote for the allele of the  
 sensitive parent (IR64)  
 10 H:    heterozygote genotype  
      B:    homozygote genotype for the allele of the  
          resistant parent (Gigante)  
      D:    genotype non homozygote for the allele of the  
          resistant parent (Gigante)  
 15 \*    under the assumption of absence of double  
      combination in space M1-R and M2-R  
      \*\*    estimated distance using resistance map on 183 F2  
          (IR64 x Gigante) cf (figure X)  
          14 bands from the resistant parent were also  
 20 identified and will or will not be confirmed on  
      recombinants generated in the F2 population (IR64 x  
      Gigante).

25 Example 13:    Anchoring of the RYMV resistance locus  
      using microsatellite markers

30        The M1 marker being positioned on chromosome 4 of  
      the genetic map (IR64 x Azucena; example 9)  
      microsatellite markers such as defined in (6) and  
      belonging to this chromosome were used to fine-tune the  
      map of the RYMV resistance locus. The following  
      microsatellite markers were tested: RM241, RM252 (1),  
      RM273 and RM177(6), under the experimental conditions  
      described in (1) and (6). With the exception of the



RM177 marker, non-polymorphous between the IR64 and *Gigante* parents, the markers RM241, RM252, RM273 were mapped on a F2 population (IR64 x *Gigante*) assessed in parallel for RYMV resistance. The results on 183 F2 individuals make it possible to characterized a stretch of approximately 3.6 cM bordered by the two microsatellite loci RM252 and RM272 surrounding the RYMV resistance gene (see figure 10(a)).

10 Example 14: Fine mapping of the space carrying the resistance locus and order of the resistance markers in the M1-M2 space.

The 45 F2 individuals (IR64 x *Gigante*) resistant and recombinant for the M1 and m2 markers were characterized for the microsatellite markers identified in example 13. The mapping of the markers in segregation on all the F2 individuals (IR64 x *Gigante*) available (321) confirms the order and the distance between the markers of the M1-M2 space, in particular the RM252-RM273 space which is estimated at 3.6 cM (figure 10(b)). With the 45 F2 individuals (IR64 x *Gigante*) that are resistant and recombinant for the M1 and M2 markers, it is possible to confirm the order of the AFLP markers identified in example 12. One AFLP marker, EACG/MACA, remains within the RM252-RM273 space and represents the nearest marker to the RYMV resistance locus (Table 9). Overall, out of the 321 F2 individuals tested, there are 20 individuals recombined on one side or other of the RYMV resistance locus and may advantageously be used to identify closer markers and/or for cloning the resistance gene.

Example 15: Marker-assisted resistance transfer

The markers close to the resistance locus were tested on irrigated varieties highly sensitive to the RYMV virus (var BG90-2, Bouaké189, Jaya). 3 markers (M1, RM241, RM252) show polymorphism between these 3 varieties and the *Gigante* variety, enabling the use of these markers to be considered for resistance transfer to sensitive genotypes. Experimental transfer of resistance to these varieties was made as far as the 2<sup>nd</sup> backcross. At each cross, the plants were verified for the presence of markers derived from *Gigante*, and resistance segregation was controlled by descendant tests on F2. Table 10 below summarizes results.

TABLE 10

Recurrent parent	Polymorphism / donor parent ( <i>Gigante</i> )					Generation obtained	theoretical % recurrent parent	N° of lines obtained
	M1	RM241	RM252	RM273	RM177			
BG90-2	poly	poly	poly	-	-	BC2F2	87.5	4
Bouaké	poly	poly	poly	-	-	BC2F2	87.5	1
189	poly	poly	poly	-	-	BC2F2	87.5	2
Jaya	poly	poly	poly	poly	mono	BC3	93.7	5
IR64								

Example 16: anchoring of the resistance gene on the physical map

The AFLP band corresponding to the M3 marker and amplified in the susceptible parent IR64 with the primers E-ACG/M-ACA has been cloned using the same conditions than for M1 marker. This band was sequenced:

Sequence of M3 marker (excluded adaptators):

Acggacctatccacttttatgccagcaagaaaatttagatgatggcaactgtatgt (seq. N°13)

DNA from varieties *Gigante*, IR64, Azucena and Tog5681 was digested using restriction enzymes Hind III, Eco RV, Dra I, Xba I, Bgl II, Bam HI, Sca I et Eco RI and membranes has been realized. Hybridization of the M3 sequence on these membranes did not reveal polymorphism

between tested varieties. However, hybridization profile revealed that M3 is a single copy sequence in rice genome. This probe has been used to screen a BAC library including 36000 clones, realized in Clemson University using DNA of Nipponbare variety, digested with Hind III enzyme.

Membrane prehybridization was performed one night at 65°C in hybridization tubes, in a buffer made of SDS 7%, sodium phosphate 0.5M pH7.2, EDTA 1 mM, salmon sperm DNA (0.1 mg/ml). Hybridization was performed in the same buffer in which labeled probe was added. Probe was radioactively labeled using the "5'-end-labelling" kit from Amersham-Pharmacia, as recommended by furnisher. After one night at 65°C, membranes were washed twice 20 minutes in SSC 1X, SDS 0.1% and twice 20 minutes in SSC 0.5X, SDS 0.1%. The, membranes were wrapped in Saran-wrap and kept at -80°C in contact with film.

Probe corresponding to M3 marker hybridized on 17 clones, 13 of which belong to contig 89, as described on Clemson University web site () on 06/12/01. These clones were : OSJNBa0006L19, OSJNBa0015F04, OSJNBa0022O14, OSJNBa0032M10, OSJNB a0048E10, OSJNBa0043I12, OSJNBa0051M11, OSJNBa0052K13, OSJNBa0059I01, OSJNBa0058F05, OSJNBa0070I17, OSJNBa0083D09. OSJNBa0087J22. These results are coherent enough to consider that M3 is on contig 89. A figure of the contig 89 is given (fig 11), clones hybridizing with M3 are indicated using a thick trait.

Several BAC of contig 89 have been sequenced in the international rice genome sequencing project and sequenced were released on data banks. Thus, the BAC clone OSJNBa0014K14, localized, at one extremity of contig 89, has been sequenced and its sequence has been recorded under accession number AL606604 on GenBank. A microsatellite sequence was identified on this clone and primers have been designed on both sides of this sequence in order to develop a microsatellite marker (referred later as MS606604-2).

**Gcaaagtgtttcaccttggacc**catgcattcctcctctctctctctctctctctctctctct  
TCTgagc  
gctcaactctccattgagcactg**agcaggcccttacctttgctt**

20 Agcaggcccttacctttgcct (sequence NB 15)

25	dNTP	200 $\mu$ M
	Taq	0.02 U/ $\mu$ l
	Buffer	1X
	MgCl <sub>2</sub>	1.5 mM
	Forward primer-M13	0.1 $\mu$ M
30	Reverse primer	0.1 $\mu$ M
	Primer M13-IRD700	0.06 $\mu$ M
	(amplification in 15 $\mu$ l)	

35 In order to visualize amplification products on a LICOR  
sequencer, amplification is performed using the M13-  
forward universal primer labeled with IRD700 and the  
forward MS606604-2 primer to which the sequence 13-  
forward is added in 5' position (tailing protocol  
described by furnisher). Amplification is realized with  
40 the program:

5 min 94°C  
 30 s 94°C  
 30 s 57°C  
 5 40s 72°C  
 (34 cycles)  
 5 min 72°C

10 A size-based polymorphism was detected between IR64 and Gigante varieties. This marker has been tested on 30 individuals recombined between RM252 and RM273 (12 resistant plants already presented in table 9 and 18 additional individuals evaluated for resistance level on F3 progenies). The marker MS606604-2 showed a  
 15 perfect co-segregation with RM252 (table 11)

Table 11

	M1	RM241	MS606604 -2	RM252	RYMV resistance	M7	RM273	M2
<u>Resistant F2 plants recombined between RM252 and RM273</u>								
F2-R17	B	-	B	B	<b>B</b>	D	H	D
F2-R20	B	B	B	B	<b>B</b>	D	H	D
F2-R25	H	-		H	<b>B</b>	B	B	B
F2-R36	H	H	H	H	<b>B</b>	D	H	D
F2-R37	H	-	H	H	<b>B</b>	B	B	B
F2-R38	B	-	B	B	<b>B</b>	D	H	D
F2-R55	H	-		H	<b>B</b>	B	B	B
F2-R61	H	-	H	H	<b>B</b>	B	B	B
F2-R93	B	B	B	B	<b>B</b>	D	H	D
F2-R105	B	B	B	B	<b>B</b>	D	H	D
F2-R135	H	H	H	H	<b>B</b>	B	H	D
F2-R141	H	-	H	H	<b>B</b>	B	B	B
<u>F2 plants recombined between RM252 and RM273, and evaluated for resistance on F3 progenies</u>								
BR5(11)	H	H		H	<b>B</b>	B	B	B
F2-1	H	H	H	H	<b>H</b>	B	B	B
F2-16	H	H	A	A	<b>A</b>	D	H	D
F2-19	H	B	B	B	<b>H</b>	-	H	D
F2-95	A	A	A	A	<b>H</b>	D	H	D
F2-113	H	-	H	H	<b>H</b>	B	B	B
F2-114	-	H	H	H	<b>B</b>	-	H	
F2-133	H	A	A	A	<b>H</b>	D	H	D

F2-142	H	-	H	H	<b>B</b>	B	B	B
F2-163	B	-	B	B	<b>B</b>	D	H	D
F2-167	B	-	H	H	<b>H</b>	D	A	D
F2-176	A	A	A	A	<b>A</b>	D	H	D
F2-184	-	-	H	H	<b>H</b>	D	A	D
F2-189	H	-	H	H	<b>H</b>	D	B	B
F2-206	-	H	H	H	<b>H</b>	B	B	
F2-223	-	A	A	A	<b>H</b>	D	H	
F2-278	B	-	B	B	<b>B</b>	-	H	D
F2-280	H	A	A	A	<b>A</b>	D	H	D
F2-285	-	-		A	<b>H</b>	-	H	

A : genotype homozygous for the allele from susceptible parent (IR64)

H: genotype heterozygous

5 B: genotype homozygous for the allele from the resistant parent (Gigante)

D: genotype not homozygous for the allele from the resistant parent (Gigante)

10 Resistance gene is localized between markers M3 and MS606604-2 and thus between the position delimited by these markers on contig 89, as mentioned on figure 11.

Bibliographic References

- (1) Chen, X et al., (1997), Development of a  
microsatellite framework map providing genome-wide  
5 coverage in rice (*Oryza sativa* L) *Theor Appl Genet* 95:  
553-567.
- (2) Panaud, O. et al., (1996), Development of  
microsatellite markers and characterization of simple  
10 sequence length polymorphism (SSLP) in rice (*Oryza*  
*sativa* L) *Mol Gen Genet* 252: 597-607.
- (3) Wu K.S. et al., (1993), Abundance, polymorphism  
and genetic mapping of microsatellites in rice. *Mol Gen*  
15 *Genet* 241: 225-235.
- (4) Zabeau et al., (1993), Selective restriction  
fragment amplification: a general method for DNA  
fingerprinting. EP 92402629.7.  
20
- (5) Vos et al., (1995), AFLP, a new technique for DNA  
fingerprinting. *Nucleic Acids research* 23: 4407-4414.
- (6) Temnyck et al., (2000), *Theor Appl Genet* 100:697-  
25 712.

The entire content of all reference cited or referred  
to herein is incorporated herein by reference.